

FINGERPRINTING AND DETECTING OF CLOSELY RELATED CUCUMBER (CUCUMIS SATIVUS L.) GENOTYPES USING APPLICATION OF ISSR MARKERS

Nidhal Abdul Hussein Al–Bdairi¹ and Balgees Hadi Al-Musawi²

¹Department of Biology, College of Education for girls, Kufa University, Iraq. ²Department of Biology, College of Science, Kerbala University, Iraq.

Abstract

This research was conducted in molecular laboratory of college of education for girls-Kufa university. ISSR (Inter Simple Sequence Repeat) markers were used to evaluate the genetic relationships among six genotypes of cucumber (*Cucumis sativus* L.) cultivated in Iraq. This study was conducted for fingerprinting ten Iraqi cucumber genotypes using ten ISSR markers; products of ISSR markers amplification produced 67 polymorphic bands out of 106. The ISSR primers UBC-807, UBC-835 and UBC-872 success in giving a unique fingerprint for most studied genotypes. The results revealed that ISSR markers could be efficiently applied to quickly access for genetic variation available in the cucumber germplasm. The information on genetic similarity among genotypes will be useful for selecting the accessions to establish a germplasm bank of cucumber landraces and to develop breeding programs.

Key words: Cucumber, ISSR, Fingerprint, Genetic diversity.

Introduction

Cucumber (*Cucumis sativus* L.) considered is an significant vegetable crop grown and a widely cultivated plant in worldwide, which belongs to the gourd Cucurbitaceae family that consist of several crops of economic value, many medicinal and nutritional benefits (Gill and Bali, 2011). Cucumber is one of important and desirable economic crops in Iraq as its fruits are used as aperitif, consumed freshly or with salad or pickles, but is the fourth extreme useful vegetable crop existence sold in the world (FAO, 2016).

Cucumis genus consist of further than 55 species universally included 25 Asian and Australian and sacrifially 30 African species as detected by DNA sequences of plastid and nuclear markers, then have two species of cucumber (*Cucumis sativus*) and melon, muskmelon (*Cucumis melon*) that are economical significant vegetable crops in numerous regions particularly wherever the cucumber is most used fresh (Dhaliwal, 2017).

China, India, Russia and USA respectively regarded are the maximum producers of cucumber (Food and Drug Administration, 2016). Cucumber (*Cucumis sativus* L.) have a small chromosome complements with n = x = 7, 2n = 2x = 14 and a little haploid genome of 367 Mbp/C. Also it is a perfect model organism for experiences which cultivated for genetic studies, because of characteristic and it is various regulate of unisexual or bisexual flowering sex phenotypes (Nam, *et al.*, 2005).

Molecular markers considered a good tools applied to recognize genetic variation in several organisms (Irshad and Idrees, 2014). DNA fingerprinting and assessment of genetic relationships have become an significant tool to varietal identity in plant breeding programs (McGregor, 2000; Wünsch and Hormaza, 2002). ISSR is one of types of DNA markers that depend on the PCR technique and due to its efficiency to recognize individuals with major accuracy and regulator (Nadeem *et al.*, 2018).

ISSR have been used to successfully to assess variation in a vast range of plants, such as evaluate genetic variety in most plant species, included a Potato (Rocha *et al.*, 2010), Barley (Guasmi *et al.*, 2012), Walnut (Ahmed *et al.*, 2012), in Ocimum species (Chen *et al.*,

*Author for correspondence : E-mail : nidhal.albdairi@uokufa.edu.iq, balqees.hadi@uokerbala.edu.iq

2013) Rice (Girma *et al.*, 2013) and *Cucurbita pepo* (Nontuthuko *et al.*, 2015).

ISSR was described as a powerful technique to assess genetic diversity and similarities between and within species. The aims of this study was evaluation of ISSR marker ability to assess genetic diversity between Iraqi cucumber genotypes in order to fingerprint and estimate genetic relationships between them. Also, results can generate invaluable basic reference or more efficient germplasm management and for origination utilized for cucumber a breeding program, such as genotypes identification, parental choice for developing heterotic hybrids or germplasm preservation.

Materials and Methods

Preparation of plant material and genomic DNA isolation

Plant material consisted of six genotypes of Cucumber (*Cucumis sativus* L.) genotype seeds were selected from different origin and certified sources in the country, such as: 1 (Rami - local), 2 (Tender green- USA), 3 (Straight eight- USA), 4 (Bush champion- USA), 5 (Spacer master - USA) and 6 (Stimora- USA).

Genome DNA Isolation

Table 1: Primers and their sequences used as ISSRs markers.

Primer	Sequence	Ann-	Primer	Sequence	Ann-
Name	5'-3'	ealing	name	5'-3'	ealing
UBC-807	$(AG)_8T$	50C ⁰	UBC-828	(TG) ₈ A	50C ⁰
UBC-808	(AG) ₈ C	52C ⁰	UBC-835	(AG) ₈ YC	48C ⁰
UBC-810	$(GA)_8T$	53C ⁰	UBC-848	(CA) ₈ RG	52C ⁰
UBC-816	$(CA)_{8}T$	52C ⁰	UBC-872	(GATA) ₄	48C ⁰
UBC-817	$(CA)_8A$	52C ⁰	UBC-881	GGG (TG	50C ⁰
				GGG)2TG	

DNA was extracted using Genomic DNA Mini Kit from the fresh studied cucumber genotypes leaves and DNA quantity, about 132 μ g/ml with a purity ranged between 1.8–1.9 which measured by Nanodrop device.

ISSR Application

In this study, using ten ISSR markers with temperature detailed in table 1. PCR amplification conducted using Thermo cycler (Agilent technology sure cycler 8800) in PCR pre mix master mix containing (250 μ M of each dNTP, 1Unit of *Taq* DNA polymerase and 1X reaction buffer, with 1.5mM MgCl₂). Then, it was placed in the Thermo-cycler on a specific program for each primers according to annealing temperatures of primer illustrated in details from table 1, According (Golabadi *et al.*, 2012). Electrophoresis done using 1.5% agarose gel at 70 Volts for 3 hours.

Statistical Analysis

The results of the amplification process gathered in a table depending on the Scoring data as 1, 0 for presence or absence of DNA bands in the studied samples, the number 1 indicates to the presence of clear DNA band, while the number 0 indicates absence of the band. Individually the tables were organized for each pimer and the dendrogram was drawn by applying UPGMA (Unweighted Pair Group Method with Arithmetic Averaging) utilize the Statistical PAST program, Version 62.1. (Hammer *et al.*, 2001).

Results and Discussion

Fingerprinting analysis

DNA fingerprinting with the objective of genotypes or varietal identification, has become an significant tool for genetic identification, germplasm management and

 Table 2: Summary of primers amplification and discrimination value of each ISSR primer in this study.
 recording systems (Wu et al., 2010). Ten primers were tested showing varied

No.	Primer name	No. of ampl- ified	No. of main bands	No. of Mono- morphic	No. of Poly- morphic	Poly -morp hism	Pri- mer effic-	Discr- -imin- -atory
		bands	Danus	Bands	bands	(%)	iency	value(%)
1	UBC-807	57	14	4	10	71.43	0.175	14.925
2	UBC-808	44	10	3	7	70	0.159	10.448
3	UBC-810	50	8	4	4*	50	0.080	5.970*
4	UBC-816	32*	6*	2*	4*	66.67	0.125	5.970*
5	UBC-817	43	9	3	6	66.67	0.140	8.955
6	UBC-828	55	11	5	6	54.55	0.110	8.955
7	UBC-835	39	9	2*	7	77.78*	0.179	10.448
8	UBC-848	53	13	9*	4*	30.77*	0.075*	5.970*
9	UBC-872	63*	17*	4	13*	76.47	0.206*	19.403*
10	UBC-881	41	9	3	6	66.67	0.146	8.955
	Total	477	106	39	67			

primers were tested showing varied multiplication results between the studied categories.

These primers showed 477 amplified bands from the original 106 main bands and the high number of amplified bands was 63 obtained through all genomes by the UBC-872 primer while the lowest number of the amplified bands was 32 by the UBC-816 primer. Also the highest number of main bands 17 was obtained by the UBC-872 primer whereas, the lowest number of main bands 6 by the UBC-816 primers.

The difference in the number of main

and amplified bands is mainly due to the primer structure and that some primers recognize a large number of link locations which are more useful than those recognizing a lower number of these locations, giving better chance of detecting DNA polymorphisms among individuals (Williams *et al.*, 1990 and Tahir, 2014). The results suggest that the UBC-872 primer should be used in the future and other races for the two reasons table 2.

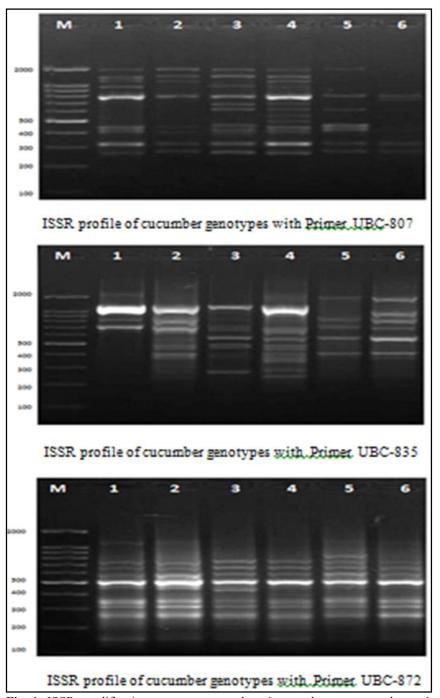


Fig. 1: ISSR amplification pattern generated on 6 cucumber genotypes lanes: 1 (Rami - local), 2 (Tender green- USA), 3 (Straight eight- USA), 4 (Bush champion- USA), 5 (Spacer master USA) and 6 (Stimora- USA). M = 100-2000 bp DNA ladder marker.

In this study, the ISSR primers (UBC-807, UBC-835 and UBC-872) that showed specified fingerprints can be applied in among to screen lengthy spectrum of cucumber access. That alleles number are significant for they likely impress for specific areas with a genome specified to a special species of cucumber by ISSR markers, Fig. 1. Also, accession on particular DNA fingerprints will be of rise value into cucumber breeders

cooperative in the improvement of the yield.

The primer UBC-835 gave higher value for polymorphism 77.78%, while the lowest value 30.77% for polymorphism in primer UBC-848 table 2, that showed a more polymorphism than the ISSR markers may provide better than genetic patterns (Demir *et al.*, 2010). The most significant fact to be possessed into account is that difference in the level of polymorphism can be the score of discrete areas of the cucumber genome that have been evaluated by select indicators or variance among the materials utilized (Sun *et al.*, 2001).

The number of primers required to identify selected varieties is dependent on discrimination power of each tested primers which means its ability to finding unique pattern for the studied genotypes. Discrimination power of a primer will increased by increasing the number of specific genotypes using the specific primers (Arif et al., 2010) It is useful to test several primers to identify the genotypes obtained. Higher discriminatory in primer UBC-872 of 19.403 while, primers UBC-810, UBC-816 and UBC-848 gave the lowest discriminating capacity of 5.970.

Also the other important criteria to be analyzed, the efficiency of the primer and Discriminatory capacity determined by the materials and methods of all the primers tested in this study. It was found that the UBC-872 primer gave the highest calculated efficiency of 0.206 while, the UBC-848 primer gave the least efficiency Calculated as 0.075 table 2.

The primer efficiency range may show the primer's ability to give a large

Table 3: The genetic distance values among 6 cucumber
genotypes: 1) Rami - local(, 2) Tender green- USA), 3
(Straight eight- USA), 4 (Bush champion- USA), 5
(Spacer master - USA) and 6 (Stimora- USA).

00000 48525 44778 39086 56393	0.00000 0.34324 0.28677	0.00000							
48525 44778 39086	0.34324								
44778 39086	0.34324								
39086									
	0.28677	0 0 40 55							
56393		0.24255	0.00000						
	0.49794	0.40650	0.26674	0.00000					
58950	0.33564	0.39914	0.33472	0.33426	0.00000				
Distance									
0.42	0.36	0.24	0.18	0.12	0.00-				
1		-L - L	<u>.</u>	I					
					2				
		Г			3				
	2								
	_				4				
					5				
					6				
					1				
	58950 045 		Distance	Distance	Distance				

Fig. 2: UPGMA dendrogram of genetic relationship tree between cucumber genotypes: 1 (Rami - local), 2 (Tender green- USA), 3 (Straight eight- USA), 4 (Bush champion- USA), 5 (Spacer master - USA) and 6 (Stimora-USA). based on ISSR data.

proportion of the polymorphic bands according to a number of multiplication ranges. Thus, the primer efficiency is not the primer which gives the largest number of multiplied packs, but the ability to show differences between the studied species (Newton and Graham, 1997). Power discrimination was also used to reveal the fingerprinting of other plants genomes such as cucumber (Al-Jaf and AL-Jubouri, 2017) and Rice (Al-Musawi *et al.*, 2019). Discrimination power reflects genetics and is very important for plant breeders to see large variation for selection in future studies.

Relationship among cucumber genotypes using ISSR markers

Based on the estimated genetic similarity matrix table 3, Value of the highest genetic distance was 0.58950 observed between 1 (Rami - local) and 6 (Stimora- USA) genotypes (both originated from different locations. While the lowest genetic distance was 0.24255 observed between 3 (Straight eight- USA), 4 (Bush champion-USA) genotypes (originated from USA), these relations between genotypes which dependent to geographical origin and related to that they share a common parent or ancestor, genetic similarity, the presence of common ancestors or not might have influence the similarity or dissimilarity among the genotypes (Morales *et al.*, 2011).

This variation between genotypes in agronomic characteristic are perhaps due to the variance of climate types, ground sample and the position to another (Ezzat *et al.*, 2010). Also, this possibly due to the effect of varies ecological conditions on the phenotypic characteristic, essential to clear variation even between similar genotypes (Shehzad *et al.*, 2009). Also the domestic names cannot be looked perfect leader to the appearance of diversity (Chakauya *et al.*, 2006).

Results in Fig. 2 show that cucumber genotypes divided between two major cluster, the first small one included only 1 (Rami) (local from Najaf), the other large major cluster contained the rest five genotypes which further divided in to two sub clusters, the first small contained two genotype while the other large sub cluster contained three genotypes.

The introductions include cucumber from USA, Therefore, the high levels of genetic variability detected in the studied collection are justified by the multiple origin of the genotypes associated with cultivation in different localities for several years and they can be applied for pedigree analysis and to define phylogenetic relationships among genotypes (Senior *et al.*, 1998). Molecular markers are therefore, important tools for preservation and for management of the existing genetic variability.

In this study, ISSR primers as announced here so, collect helpful information for evaluating the genetic diversity among cucumber germplasm in Al-Najaf areas and for the use of cucumber landraces in Iraq in future breeding of new genotypes of cucumber that may be most tolerant to abiotic stresses, more productive and have better nutritive value contrast to the present landraces.

Conclusion

Molecular markers (ISSR) powerful and useful tool for clarifying relationships within species and also for providing useful genetic indicators to identify genotypes in the cucumber and polymorphisms found among species that can be used in breeding programs to maximize the use of genetic resources. Also that are a further reliable and careful method for diversity estimation.

The information acquired from this study possibly

helpful for then identification of favorable cucumber genotypes for realization the reach of genetic diversity current in cucumber genotypes.

References

- Ahmed, N., J.I. Mir and R.R. Mir (2012). SSR and RAPD analysis of genetic diversity in walnut (*Juglans regia* L.) genotypes from Jammu and Kashmir, India. *Physiol. Mol. Biol. Plants*, 18: 149-160.
- Al-Jaf, I.H. and K.A. AL-Jubouri (2017). Determination of Genetic Distance among Genotypes of Cucumber, *Cucumis* sativus L., Using ISSR Marker. Journal of Agriculture and Veterinary Science, 10(4): 33-37.
- Al-Musawi, B.H., N.A. Al-Bdairi and M.A. Al-Anbari (2019). Molecular Characterization Based on Genetic Diversity Analysis For Fertility Restorer Genes in Rice (*Oryza sativa* L.) Using Microsatellite Markers. *International Journal* of Pharmaceutical Research, 11(2): 81-89.
- AL-Musawi, B.H. (2019). Morphological, Cytological and Molecular Study on many Rice (*Oryza sativa* L.) Genotypes. Ph.D. thesis College of Education for pure sciences, University of kerbala, Iraq.
- Arif, I.A., M.A. Bakir, H.A. Khan, A.H. Al-Farhan, A.A. Al-Homaidan, A.H. Bahkali, M. Al-Sadoon and M. Shobrak (2010). Application of RAPD for molecular characterization of plant species of medicinal value from an arid environment. *Genet. Mol. Res.*, 9(4): 2191-2198.
- Chakauya, E., P. Tongoona, E.A. Matibiri and M. Grum (2006). Genetic Diversity Assessment of Sorghum Landraces in Zimbabwe Using Microsatellites and Indigenous Local Names. *International Journal of Botany*, 2: 29-35.
- Chen, S.Y., T.X. Dai, Y.T. Chang, *et al.*, (2013). Genetic diversity among Ocimum species based on ISSR, RAPD and SRAP markers. *Aust. J. Crop. Sci.*, **7:** 1463-1471.
- Demir, K., M. Bakýr, G. Sarýkamýþ and S. Acunalp (2010). Genetic diversity of eggplant (Solanum melongena) germplasm from Turkey assessed by SSR and RAPD markers, Genetics and Molecular Research, 9(3): 1568-1576.
- Dhaliwal, M.S. (2017). Cucurbits. In: Handbook of Vegetable Crops, third ed. Kalyanis publishers. Darya Ganj, New Delhi, India, pp. 77-147.
- Ezzat, E.M., M.A. Ali and A.M. Mahmoud (2010). Agronomic Performance, Genotype X Environment Interactions and Stability Analysis of Grain Sorghum (Sorghum bicolor L. Moench). Asian Journal of Crop Science, 2: 250-260.
- FAO (2016). FAOSTAT. Food and Agriculture Organization of the United Nations, Rome, Italy.
- Food and Drug Administration (2016). Cucumbers. The Western Institute for Food Safety and Security. University of California-Davis, Davis, CA, USA.
- Gill, N.S. and M. Bali (2011). Isolation of antiulcer Cucurbitane type triterpenoid from the seeds of Cucurbitapepo.

Research Journal of Phytochemistry, 5(2): 70-79.

- Girma, G., K. Tesfaye and E. Bekele (2013). Inter Simple Sequence Repeat (ISSR) analysis of wild and cultivated rice species from Ethiopia. *Afr. J. Biotechnol.*, **9:** 5048-5059.
- Golabadi, M., P. Golkar and A. Eghtedary (2012). Assessment of genetic variation in cucumber (*Cucumis sativus* L.) genotypes. *European Journal of Experimental Biology*, 2(5): 1382-1388.
- Guasmi, F., W. Elfalleh, H. Hannachi, K. Feres, L. Touil, N. Marzougui, T. Triki and A. Ferchichi (2012). The use of ISSR and RAPD markers for genetic diversity among South Tunisian barley. ISRN Agronomy, 1-10.
- Hammer, D., A. Harper and P. Ryan (2001). PAST : Paleontological Statistics.
- Irshad, M. and M. Idrees (2014). Molecular markers in plants for analysis of genetic diversity: a review. *Eur. Acad. Res.*, 2: 1513-1540.
- McGregor, C.E., C.A. Lambert, M.M. Greyling, J.H. Louw and L. Warnich (2000). Acomparative assessment of DNA fingerprinting techniques (RAPD, ISSR, AFLP and SSR) in tetraploid potato (*Solanum tuberosum* L.) germplasm. *Euphytica*, **113**: 135-144.
- Morales, M., V. Decker and L. Ornella (2011). Analysis of genetic diversity in Argentinian heterotic maize populations using molecular markers. *Cien. Inv. Agric.*, **37(1):** 151-160.
- Nadeem, M.A., M.A. Nawaz and M.Q. Shahid (2018). DNA molecular markers in plant breeding: current status and recent advancements in genomic selection and genome editing. *Biotechnol. Biotechnol. Equip.*, **32**: 261-285.
- Nam, Y.W., J.R. Lee, K.H. Song, M.K. Lee, M.D. Robbins, S.M. Chung, J.E. Staub and H.B. Zhang (2005). Construction of two BAC libraries from cucumber (*Cucumis sativus L.*) and identification of clones linked to yield component quantitative trait loci. *Theor Appl Genet.*, **111**: 150–161.
- Newton, C.R. and A. Graham (1997). Polymerase Chain Reaction. 2nd ed. Bios. Scientific Publishers Ltd., Oxford, U.K.
- Nontuthuko, R.N., B.T. Pangirayi and M.Z. Alpheus (2015). Genetic diversity in landraces revealed by RAPD and SSR markers. *Sci. Hortic.*, **189:** 192-200.
- Rocha, E.A., L.V. Paiva, H.H.D. Carvalho and C.T. Guimaraes (2010). Molecular characterization and genetic diversity of potato cultivars using SSR and RAPD markers. *Crop Breed. Appl. Biotechnol.*, **10**: 204-210.
- Senior, M.L., J.P. Murphy, M.M. Goodman and C.W. Stuber (1998). Utility of SSRs for determining genetic similarities and relationships in maize using an agarose gel system. *Crop Sci.*, **38**: 1088-1098.
- Shehzad, T., H. Okuizumi, M. Kawase and K. Okuno (2009). Development of SSR-based sorghum (*Sorghum bicolor* L. Moench) diversity research set of germplasm and its evaluation by morphological traits. *Gen. Res. Crop Evol.*, 56: 809-827.
- Sun, G.L., M. William, J. Liu, K.J. Kasha and K.B. Pauls (2001).

Microsatellite and RAPD polymosphisms in Ontario corn hybrids are related to the commercial sources and maturity ratings. *Mol. Breed.*, **7:** 13-24.

- Tahir, N.A. (2014). Genetic Variability Evaluation Among Iraqi Rice (*Oryza sativa* L.) Varieties using RAPD Markers and Protein Profiling. *Jordan Journal of Biological Sciences*, 7(1): 13–18.
- Williams, J.G., A.R. Kubelik, K.J. Livak, J.A. Rafalski and S.V. Tingey (1990). DNA polymorphisms amplified by arbitrary

primers are useful as genetic markers. *Nucleic Acids Res.*, **18:** 6531-6535.

- Wu, Y.G, Q.S. Guo, J.C. He, Y.F. Lin, L.J. Luo and G.D. Liu (2010). Genetic diversity analysis among and within populations of Pogostemon cablin from China with ISSR and SRAP markers. *Biochem. Syst. Ecol.*, **38**: 63-72.
- Wünsch, A. and J.I. Hormaza (2002). Cultivar identification and genetic fingerprinting of temperate fruit tree species using DNA markers. *Euphytica*, **125**: 59-67.